Cryopreservation and recovery of sperm Cryopreservation & Assisted Reproduction Laboratory Frederick National Laboratory for Cancer Research (FNL)

Sperm cryopreservation: As previously described (Ostermeier et al, 2008), a cryoprotective media (CPM) containing 18% (w/v) Raffinose (Sigma Aldrich; cat # R7630) and 3% (w/v) skim milk (BD Diagnostics; cat # 232100) is clarified, filtered and then supplemented with 477 μM alpha-monothioglycerol (MTG; FW=108.16; D=1.295g/mL; Sigma Aldrich; cat # M6145). Two mL of the CPM is used to collect the sperm from the epididymides and vas deferentia of two 3-to 6-month-old mutant males.

The sperm+CPM is loaded into 0.25 mL French straws (IMV; Maple Grove, MN; cat# AAA201) by first creating a 1.5 cm column of CPM, a 1.5 cm column of air and four 0.5 cm columns of sperm sample (~10 µL) each separated by an approximate 0.5 cm column of air and the last being followed by a final column of air. The straws are heat sealed and then exposed to LN₂ vapor before being stored in the liquid phase.

In vitro fertilizations (IVF): The *In vitro* fertilization medium, Research Vitro Fert (RVF; Cook Medical; Brisbane, Australia; cat# K-RVFE-50), is used for sperm incubation, IVF and zygote culture. The IVF dishes contain one 500 μ L fertilization drop and three 150 μ L wash drops, which are covered by mineral oil (Sigma cat #M8410). The dishes are equilibrated overnight in a humidified 37° C incubator maintained at 5% CO₂.

The straw containing sperm is transferred directly from LN_2 into a $37^{\circ}C$ water bath and incubated for ~30 to 60 sec. The straw is removed and lightly dried with a kim wipe. The sealed end of the straw is cut off and a stylet used to push a single 10 μL aliquot of CPM+sperm into a fertilization drop. This can be repeated up to four times per straw. The sperm are incubated in the fertilization drop for ~1

hr prior to the addition of cumulus oocyte complexes from five superovulated 17-to 27-day-old female mice. Superovulation is achieved by intraperitoneal injections of 5 IU PMSG (Calbiochem (EMD Millipore); cat # 367222) followed 48 hours later by 2.5 IU hCG (Sigma; cat # C1063).

After 4 hrs of co-incubation of the sperm and oocytes, the presumptive zygotes are washed and only those appearing normal are cultured overnight in one of the 150 μ L RVF wash drops.

Approximately 18 hrs after washing, two cell embryos are collected and transferred into B6D2F1 pseudopregnant recipients. We commonly do single sided transfers and recommend that 10-15 embryos be transferred per recipient.

References:

Ostermeier GC, Wiles MV, Farley J, Taft RA. Conserving, distributing and managing genetically modified mouse lines by sperm cryopreservation. PLoS ONE. 2008;3(7): e2792.